DOI: https://doi.org/10.56286/y2wkv643





P-ISSN: 2788-9890 E-ISSN: 2788-9904

NTU Journal of Agricultural and Veterinary Sciences





Natural Cholinesterase Activity in Some Local Animals: Review

1st Ashraf. S. Alias Department of Environmental Health /College of Environmental Sciences /University of Mosul

Article Informations

Received: 27-08- 2024, **Accepted:** 28-01-2025, **Published online:** 28-06-2025

Corresponding author:

Name: Ashraf. S. Alias Affiliation: Department of Environmental Health /College of Environmental Sciences /University of Mosul

Email:

ashraf.saddik@uomosul.edu.iq

Key Words:

Cholinesterase, Modified Electrometric Method, local animals, Insecticide, Organophosphorus.

ABSTRACT

Organophosphorus pesticides are common causes of poisoning due to their easy access to local markets, low price, lack of awareness about them, and misuse by the user, which exposes the environment surrounding animals and birds to poisoning by these pesticides. The current study aims to provide the normal values of cholinesterase enzyme in some field animals, whose activity was measured using a simple electrical method to measure blood samples (plasma and red blood cells) of animals. References related to the normal activities of cholinesterase enzyme in a number of animals were cited, including (cows, sheep, buffalo, goats, dogs, and cats), and cholinesterase enzyme activity (ChE) was measured using the electrical measurement method. The electrical measurement method was effective, simple, accurate, and reproducible to detect exposure of animals in the natural environment to organophosphate pesticides or carbamates as an indicator of environmental pollution.



©2023 NTU JOURNAL OF AGRICULTURAL AND VETERINARY SCIENCES, NORTHERN TECHNICAL UNIVERSITY. THIS IS AN OPEN ACCESS ARTICLE UNDER THE CC BY LICENSE: https://creativecommons.org/licenses/by/4.0/

Introduction

Compounds such as carbamates organophosphorus are used as pesticides in public health, farming, and veterinary medicine [1]. to control zoonotic diseases and treat internal and external parasites [2,3]. The misuse of carbamates and organophosphorus can lead to environmental contamination and toxicity through exposure [4,5,6,7,8]. Intoxication occurs through inhibition (ChE) activity activity an enzyme essential for normal nervous system function, and prolonged exposure to it can lead to neurological issues and reproductive disorders caused by causing accumulation of acetylcholine (Ach) in nerve terminals, leading to eventual death with nicotine-, muscarinic-, and central nervous system-related intoxication [9, 10]. Assessing blood ChE activity can help detect influences by organophosphorus and carbamate pesticides At the beginning of poisoning exposure. A 25-30% decrease in cholinesterase activity in exposure organophosphorus or carbamate pesticides is the first step to poisoning. Cholinesterase activity reduced by 25-30%Red blood cells or plasma indicates exposure to this toxicant [1, 6]. A number

of methods appear to detect cholinesterase activity, such as the Ellman method [10]. Michel method [11] and improved electrometric measurement method [12]. The latter method is inexpensive, measures more samples in less time, is simple, and requires many tools such as a water bath and pH meter This study aims to present the results of a modified electrometry method obtained by researchers at the University of Mosul by measuring "cholinesterase activity in red blood cells and plasma" of various animals. How about conducting the same study or using animal samples detect environmental contamination by pesticides, comparing the data with informed researchers to detect exposure or poisoning from these pesticides. These values are a starting point for future research using the values described here.

RESULTS

Table (1) shows the normal value in plasma and erythrocyte cholinesterase activity in a number of local animal species.

Table 1. Normal cholinesterase activity in local animals

Species	Sex	Cholinesterase	ΔрΗ	Base material	Incubation time/ mints	Reference
	Male	plasma	0.10	Acetylcholine iodide		
Cattle _		erythrocytes	0.91		20	[13,14]
	Female	plasma	0.19	Acetylcholine iodide	_	[13,14]
		erythrocytes	0.86			
	Male	plasma	0.22	Acetylcholine iodide		
Goats		erythrocytes	0.54		40	[15,16]
	Female	plasma	0.22	Acetylcholine iodide	_	
		erythrocytes	0.44			
	Male	plasma	0.21	Acetylcholine iodide		
Sheep		erythrocytes	0.63		30	[17,18,19]
	Female	plasma	0.19	Acetylcholine iodide	_	
		erythrocytes	0.62	-		
Cats	Male	Plasma	0.37	Acetylcholine iodide	30	
		Erythrocytes	0.63		30	[20]
Dogs	Male	Plasma	0.81	Acetylcholine iodide	30	[21]
		Erythrocytes	0.67		30	
Buffalo	Female	Plasma	0.075	Acetylcholine iodide	40	[22]
		Erythrocytes	0.46		40	
Rats	Male	Plasma	0.29	Acetylcholine iodide	30	[23]
		Erythrocytes	0.29		30	
Rabbits	Male	Erythrocytes	0.45	Acetylcholine iodide	30	[24]
Chickens	Mixed breed	Plasma	0.49	Acetylcholine iodide	30	[25]
Rock dove	Female	Plasma	1.28	Acetylthiocholine iodide	30	[26]
Sand grouse	Male	Plasma	1.81	Acetylthiocholine iodide	30	[27]
Quail	Female	Plasma	1.23	Acetylthiocholine iodide	30	[28]
Starling	Male	Plasma	1.10	Acetylthiocholine iodide	30	[29]
Local Doves	Mixed	Plasma	1.04	Acetylcholine iodide	30	[30]

shows the normal blood (plasma, erythrocyte) cholinesterase activity values of different local animal species.

Conclusion

Electrometric methods may be a practical and simple method for detecting ChE in the blood tissue of animals exposed directly or indirectly to phosphorus and carbamate pesticides. This method has proven its ability to bio-assess the environmental contamination of local animals with anticholinesterase pesticides.

References. All references must follow the example format at the end of this document, and the reference list must include all cited literature.

Acknowledgments.

We thank the Department of Environmental Health, "College of Environmental Science and Technologies, University of Mosul, Iraq" for their support and for completing this study.

References

- Wilson BW.(1998). Cholinesterase inhibition. In Encyclopedia of Toxicology Vol. 1. Wexler P, Ed. Academic Press, San Diego, CA, 326-340.
- [2] Coggon, D. (2002). Work with pesticides and organophosphate sheep dips. *Occupational Medicine*, *52*(8), 467-470. https://doi.org/10.1093/occmed/52.8.467.
- [3] Jaga, K., & Dharmani, C. (2003). Sources of exposure to and public health implications of organophosphate pesticides. *Revista panamericana de salud pública*, 14, 171-185.
 - [4] Wilson BW. (1999). Clinical enzymology. In The Clinical Chemistry of Laboratory Animals. Loeb WF, Quimby FW, Eds. Taylor and Francis, Philadelphia, PA, 399-454.
 - [5] Aygun, D., Doganay, Z., Altintop, L., Guven, H., Onar, M., Deniz, T., & Sunter, T. (2002). Serum acetylcholinesterase and prognosis of acute organophosphate poisoning. *Journal of toxicology: clinical toxicology*, 40(7), 903-910. https://doi.org/10.1081/CLT-120016962
- [6] Kwong, T. C. (2002). Organophosphate pesticides: biochemistry and clinical toxicology. *Therapeutic drug monitoring*, 24(1), 144-149.
- [7] Rusyniak, D. E., & Nañagas, K. A. (2004, June). Organophosphate poisoning. In Seminars in neurology (Vol. 24, No. 02, pp. 197-204). Copyright© 2004 by Thieme Medical Publishers, Inc., 333 Seventh Avenue, New York, NY 10001, USA
- [8] Worek, F., Koller, M., Thiermann, H., & Szinicz, L. (2005). Diagnostic aspects of organophosphate poisoning. *Toxicology*, 214(3), 182-189.https://doi.org/10.1016/j.tox.2005.06.012
- [9] Wills JH.(1972). The measurement and significance of changes in the cholinesterase activities of erythrocytes and plasma in man and animals. CRC Crit Rev Toxicol, 1: 153-202.

- [10] Ellman, G. L., Courtney, K. D., Andres, V. and Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharmacol., 7: 88 - 95.
- [11] Michel, H. O.(1949). An electrometric method for the determination of the red blood cell and plasma cholinesterase activity. J. Lab. Clin . Med., 34: 1564 - 1568.
- [12] Mohammad, F. K., & St Omer, V. E. (1982). Modifications of Michel's electrometric method for rapid measurement of blood cholinesterase activity in animals: a
 - minireview. Veterinary and human toxicology, 24(2), 119-121.
- [13] Mohammad, F. K., Bhattacharyya, H. K., Fazili, M. R., Nasreen, S., Jeelani, S. G., Sheikh, N. A., ... & Qureshi, S. (2007). Review of a Practical Electrometric method for determination-of Blood and Tissue Cholinesterase activities in Animals. feedback, 2, 16.
- [14] Mohammad, F. K. (2022). Clarifying an electrometric method for determining blood cholinesterase activity: a scientific letter. Asia Pacific Journal of Medical Toxicology, 11(1), 30-32.
- [15] Al-Jobory, M. M. H., & Mohammad, F. K. (2004). A pH method for measuring blood cholinesterase activity in goats. In Abstract book of the 12th congress of mediterranean federation for health and production of ruminants, Istanbul, Turkey (Vol. 75).
- [16] Al-Jobory, M. M., & Mohammad, F. K. (2005). Validation of an electrometric blood cholinesterase measurement in goats. *Journal of Veterinary Science*, 6(4), 299-303. DOI: https://doi.org/10.4142/jvs.2005.6.4.29.
- [17] Mohammad, F. K., Alias, A. S., Faris, G. M., & Al-Baggou, B. K. (2007). Application of an electrometric method for measurement of blood cholinesterase activities in sheep, goats and cattle treated with organophosphate insecticides. *Journal of Veterinary Medicine Series A*, 54(3), 140-143.
- [18] Mohammad, F. K., Al-Zubaidy, M. H. I., & Alias, A. S. (2007). Their in vitro Inhibition by Anticholinesterase Insecticides. *Journal of Pharmacology and Toxicology*, 2(2), 131-141. https://doi.org/10.1111/j.1439-0442.2007.00867.x
- [19] Mohammad, F. K., Al-Baggou, B. K., Alias, A. S., & Faris, G. A. (2006). Application of an electrometric method for measurement of in vitro inhibition of blood cholinesterases from sheep, goats and cattle by dichlorvos and carbaryl. VETERINARNI MEDICINA-PRAHA-, 51(2), 45.
- [20] Kata, A. A., & Alias, A. S. (2013). Application of an electrometric method for measurement of blood cholinesterase activities in cats. *Al-Anbar Journal* of *Veterinary Sciences*, 6(1), 230-241.

Ashraf. S. Alias /NTU Journal of Agricultural and Veterinary Sciences (2025) 5 (2): 140-143

- [21] Alias, A. S. (2014). Electrometric method determination of blood cholinesterase activity in stray dogs. *Iraqi Journal of Veterinary Sciences*, 28(2), 153-159. 10.33899/ijvs.2014.116934.
- [22] Alias, A. S. (2018). The use of the Electrometric Method in Measuring the Activity of Cholinesterase in Local Female Buffalo in north of Iraq. Kufa Journal For Veterinary Medical Sciences, 9(2), 13-22. https://doi.org/10.36326/kjvs/2018/v9i24169.
- [23] Kasim Mohammad, F., Abdul-Mune'm Faris, G., & Khalid Shindala, M. (2002). Comparative antidotal effects of diphenhydramine and atropine against dichlorvos-induced acute toxicosis in rats. Veterinarski arhiv, 72(1), 19-28. https://hrcak.srce.hr/78390
- [24] F. K. (1999). Reduction of dichlorvos-induced toxicosis in rabbits by levamisole. *Veterinarski arhiv*, *69*(1), 29-37.
- [25] Aldabagh, I. I., & Mohammad, F. K. (1999). Reduction of dichlorvos-induced toxicosis in rabbits by levamisole. *Veterinarski arhiv*, 69(1), 29-37.
- [26] S ALIAS, A. (2006). Effect of dichlorvos on cholinesterase activity in pigeons (rockdov). *Iraqi Journal of Veterinary Sciences*, 20(2), 191-202. DOI: 10.33899/ijvs.2006.45797.
- [27] Alias, A. S., & Mohammad, F. K. (2005). Electrometric measurement of plasma and
- [28] Alias, A., Al-Zubaidy, M., Mousa, Y., & Mohammad, F. (2011). Plasma and whole brain cholinesterase activities in three wild bird species in Mosul, IRAQ: inhibition by insecticides. *Interdisciplinary toxicology*, 4(3), 144-148.
- [29] Mohammad, F. K., & Al-Baggou, B. (2005). Electrometric cholinesterase determination in poultry treated with dichlorvos and carbaryl.
- [30] Alias, A. S. (2009). The use of an electrometric method for measurement of cholinesterase activity in plasma and tissues of local doves.